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Molecular Modeling Studies of Cholic Acid Host:

Shape And Dimensions. II.

by

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19. ABSTRACT (Continue on reverse if necessary and identify by block number)

Earlier studies Technical Report No. 3 on the three-dimensional "clam" or cup shapes formed by two cholic acid molecules linked together by a substituted aromatic spacer had shown that the resulting cavities were of similar shape but different width, due to the flexible nature of this host system, with its sensitivity to minor bond rotations. In the L-arabinose binding protein, the binding pocket shape is cup-like, approximately 12-12.3 Å long and 7.3 Å wide (taken from crystallographic coordinates, Fig. 1). In order to mimic the sugar binding proteins, wherein the sugar hydroxyl groups are hydrogen bonded to the ionized amino acids of the protein through the heteroatoms (with or without an intervening water network, Fig. 2) it was necessary to investigate the dimensions of the cholic acid host cavities more closely.

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19. ABSTRACT -continued-

The present study extends th- earlier report by defining a "standard"
for the cholic acid host structure, one that uses two dihedral angles,
each CH_2 to the aromatic ring, to define the origin direction of the
two arms (or halves) of the clam shapes. The definition
is applied to cholic acid host systems containing a m-xylylene and a
p-xylylene aromatic spacer. Subsequent manipulation of two critical
dihedral angles in the flexible portion of the hosts creates cavities
which are then evaluated geometrically by measuring the interatomic
distances between key heteroatoms (hydroxyls) in each arm. The 90
and 120 degree clam shapes of the m-xylylene cholic acid host are
the closest approximations to the L-arabinose binding protein,
although these shapes are not perfect "clams" since they have the top
arm offset relative to the bottom arm. In the p-xylylene cholic acid
hosts, two 90 degree structures seem to create good clam shapes in
which to dock a monosaccharide guest. Geometry optimization with
energy minimization of these two hosts results in very little change
in the cavity shapes but does relieve several bad steric interactions.
The size of the p-xylylene cavities are approximately 12 Å long an
9-11 Å wide.

The Cholic Acid Host System Shape

Abstract

Earlier studies on the three-dimensional "clam" or cup shapes formed by two cholic acid molecules linked together by a substituted aromatic spacer had shown that the resulting cavities were of similar shape but different width, due to the flexibile nature of this host system, with its sensitivity to minor bond rotations (see report entitled The Cholic Acid Host System Shape of 2/25/90). In the L-arabinose binding protein, the binding pocket shape is cup-like, approximately 12-12.3 Å long and 7.3 Å wide (taken from crystallographic coordinates, Fig. 1). In order to mimic the sugar binding proteins, wherein the sugar hydroxyl groups are hydrogen bonded to the ionized amino acids of the protein through the heteroatoms (with or without an intervening water network, Fig.2), it was necessary to investigate the dimensions of the cholic acid host cavities more closely.

The present study extends the earlier report by defining a "standard" for the cholic acid host structure, one that uses two dihedral angles, each CH_2 to the aromatic ring, to define the origin direction of the two arms (or halves) of the clam shapes. The definition is applied to cholic acid host systems containing a m-xylylene and a p-xylylene aromatic spacer. Subsequent manipulation of two critical dihedral angles in the flexible portion of the hosts creates cavities which are then evaluated geometrically by measuring the interatomic distances between key heteroatoms (hydroxyls) in each arm. The 90 and 120 degree clam shapes of the m-xylylene cholic acid host are the closest approximations to the L-arabinose binding protein, although these shapes are not perfect "clams" since they have the top arm offset relative to the bottom arm. In the p-xylylene cholic acid hosts, two 90 degree structures seem to create good clam shapes in which to dock a monosaccharide guest. Geometry optimization with energy minimization of these two hosts results in very little change in the cavity shapes but does relieve several bad steric interactions. The size of the p-xylylene cavities are approximately 12 Å long and 9-11 Å wide.

Meta-xylylene Aromatic Spacer

The torsional angle values for bonds labeled **a**, **b**, **c**, **d**, and **e** were obtained from crystallographic coordinates (refcode = BUGJES), while **f** and **g** were set to create the reference conformation shown below. While **a**, **c**, **d**, and **e** remained fixed in both halves of the cholic acid host containing the meta-xylylene spacer, dihedral angles **b** (C_{17}--C_{20}--C_{22}--C_{23}) and **g** (benzylic C--C_1--C_2--H_{ar}) were varied. Dihedral angle **b** has the most direct effect on the cholic acid arm vector (directionality), whereas dihedral angle **g** alleviates the bad steric interactions, found in the reference torsional angle of 0 degrees, between the hydrogen of the N-H in the amide and the ortho-proton C_2-H of the benzene ring.

Reference

a = +173.6
b = +60.6
c = +179.3
d = -168.7
e = +172.6
f = +180.0
g = 0
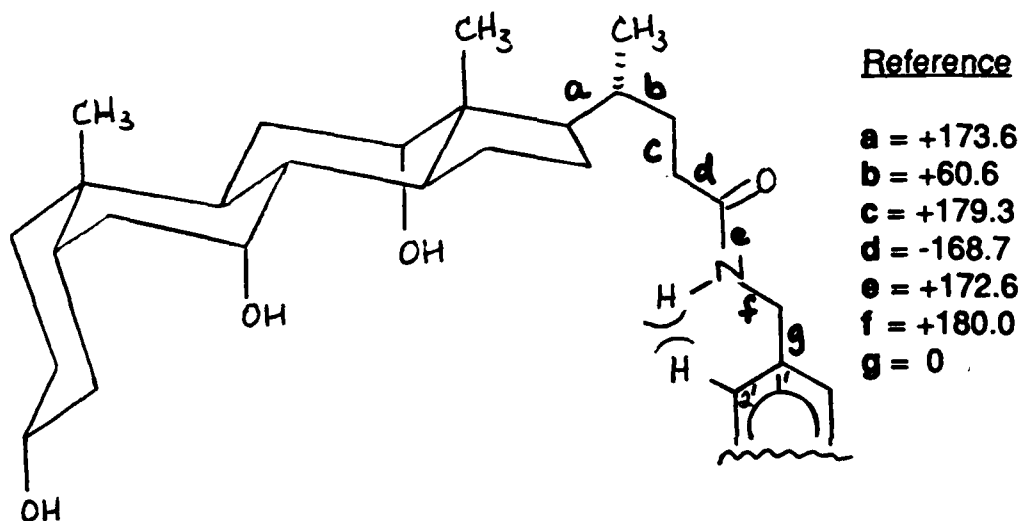
The **"standard"** for the cholic acid host is defined as shown: The molecule starts at the left, proceeding along arm **A** through the aromatic spacer toward arm **B**, always with a negative **g** dihedral angle in arm **A**, using C_1 and C_2. Dihedral angle **g** to arm **B** is defined using C_3 and C_2.

Arm A -
g defined as
N-C-C_1-C_2 NEGATIVE

Arm B -
g defined as
N-C-C_3-C_2
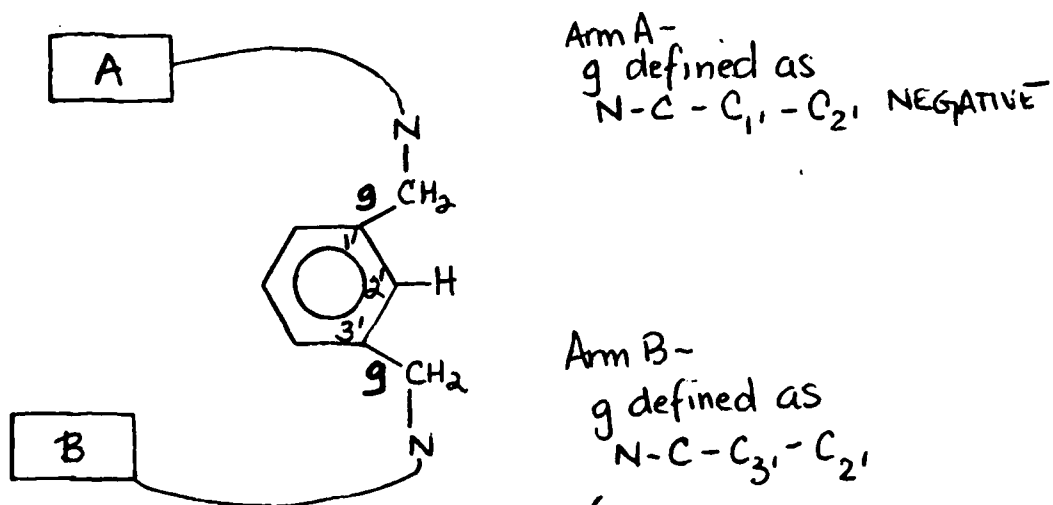
The following table of dihedral angle combinations thus results from the manipulation of dihedral angle **g**.

	Dihedral Angles			
Name	Bond 20-22 (**b**)		Bond **g** to Aromatic Ring	
	Arm A	**Arm B**	**Arm A**	**Arm B**
_60 Clam	+60.6	+60.6	-60	+60
_90 Clam	+60.6	+60.6	-90	+90
_120 Clam	+60.6	+60.6	-120	+120
_150 Clam	+60.6	+60.6	-150	+150

Each of the four cholic acid hosts above have the top arm **A** offset relative to the bottom arm **B**, thereby creating the effect of hinge movement. Although the general shape is that of a "C" or clam, the cavities are of different dimensions (Fig. 3). The interatomic distances (in Å) between the cholic acid hydroxyls in each arm reflect the cavity size, which differ mainly in width.

Name	**Å Distance**	**Arm A // B**	**O3**	**O7**	**O12**
		O3	7.31	7.73	5.18
_6C Clam		O7	10.12	10.09	5.81
		O12	7.31	7.75	3.97

		A // B	**O3**	**O7**	**O12**
		O3	8.66	10.87	7.51
_90 Clam		O7	11.98	12.54	8.23
		O12	10.64	11.48	7.84

		A // B	**O3**	**O7**	**O12**
		O3	13.77	15.75	12.11
_120 Clam		O7	15.72	16.21	12.08
		O12	14.85	15.28	11.86

		A // B	**O3**	**O7**	**O12**
		O3	18.44	19.67	16.34
_150 Clam		O7	19.38	19.38	15.77
		O12	17.97	17.77	14.91

Comparing the above synthetic cavities to the dimensions of L-arabinose binding protein, 12 Å long and 7Å wide, the _60 Clam is clearly too small in the interior. In addition, this clam shape has its ends (A-rings of the two steroidal arms) overlapping. The _150 Clam shape clearly has the largest cavity, although this may indeed be too large for a monosaccharide to maximize its hydrogen bonding interactions with both steroidal arms. The _120 and _150 Clam structures, however, seem to be reasonable

cavities in which to dock a sugar guest , with the _90 Clam most closely resembling the cavity in L-arabinose binding protein.

Manipulation of dihedral angles **b** (C_{20}-C_{22}) and **g** were done in several structures to evaluate the effect on the cavity accompanying changes in the two most crucial bonds of the flexible chain of each cholic acid arm. The following table shows the single modifications which were made.
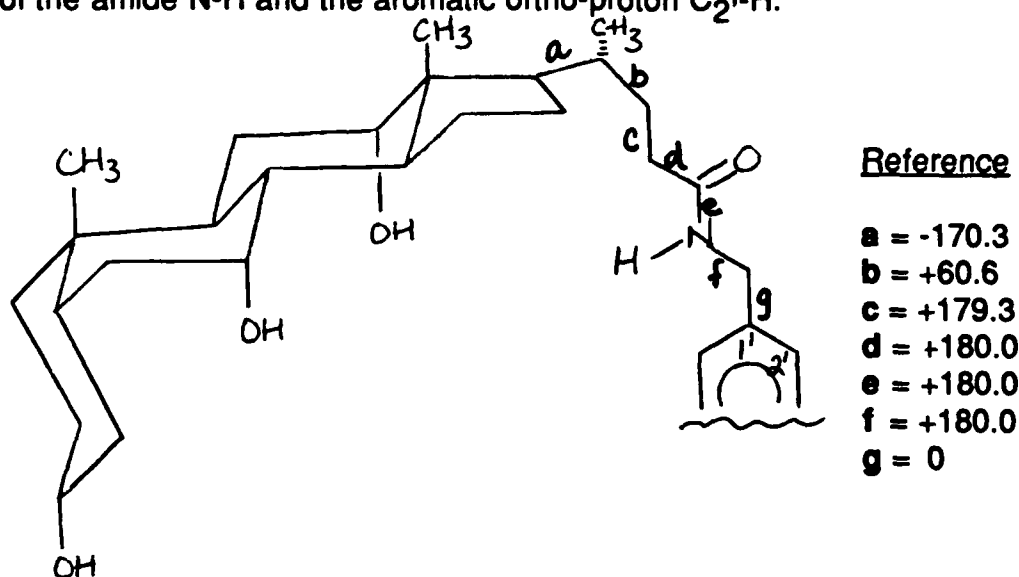
	Dihedral Angles			
Name	Bond 20-22 (b)		Bond g to Aromatic Ring	
	Arm A	Arm B	Arm A	Arm B
_90b	-60.6	+60.6	-90	+90
_90bE	-60.6	+60.6	+90	+90
_90E	+60.6	+60.6	+90	+90
_120E	+60.6	+60.6	-120	-120
_150MI*	-60.6	-60.6	+150	-150

* the total mirror image, created by reflection of X-coordinates through YZ plane

The _90b has a negative dihedral angle **b** and does not create a clam shape since the hydroxyls on arm **A** are directed outward instead of inward (Fig. 4). Structure _90bE, the extended form of _90b, and _90E, _120E, the extended forms of _90 Clam and _120 Clam respectively, illustrate that an open conformation (as opposed to a folded conformation) can be created with a single dihedral angle modification, namely **g** in arm **A** or **B** (Fig. 5, _90Clam and _90E). The total mirror image of one conformation, the _150 Clam, shows the same size cavity but opposite orientation in space (Fig. 6). None of these alternate conformations wll be considered as candidates for docking the sugar guests.

Para-xylylene Aromatic Spacer

As with the m-xylylene spacer, the torsional angles for bonds labeled **a**, **b**, **c**, **d**, and **e** were obtained from crystallographic coordinates (refcode = CUBSUN for **a**, modification of others to mimic BUGJES), while **f** and **g** were set to create the reference conformation shown below. Dihedral angles **a**, **c**, **d**, **e**, and **f** remained fixed in both halves of the cholic acid host, while **b** (C_{17}--C_{20}--C_{22}--C_{23}) and **g** (benzylic C--C_1---C_2---H_{ar}) were varied. Again, dihedral angle **b** has the most direct effect on the cholic acid arm vector (directionality), whereas dihedral angle **g** alleviates the bad steric interactions, found in the reference torsional angle of 0 degrees, between the hydrogen of the amide N-H and the aromatic ortho-proton C_2-H.

Reference

a = -170.3
b = +60.6
c = +179.3
d = +180.0
e = +180.0
f = +180.0
g = 0

The "standard" for this cholic acid host is defined as shown: The molecule starts at the left and proceeds along arm **A** through the aromatic spacer toward arm **B**, always with a negative **g** dihedral angle in arm **A**, using C1' and C2'. This dihedral angle is comparable to that used in the "standard" for the m-xylylene spacer. Dihedral angle **g** in arm **B** is measured using C_4' and C_3'.

Arm A ~
g defined as
N-C-$C_{1'}$-$C_{2'}$ NEGATIVE

Arm B ~
g defined as
N-C-C_4'-C_3'
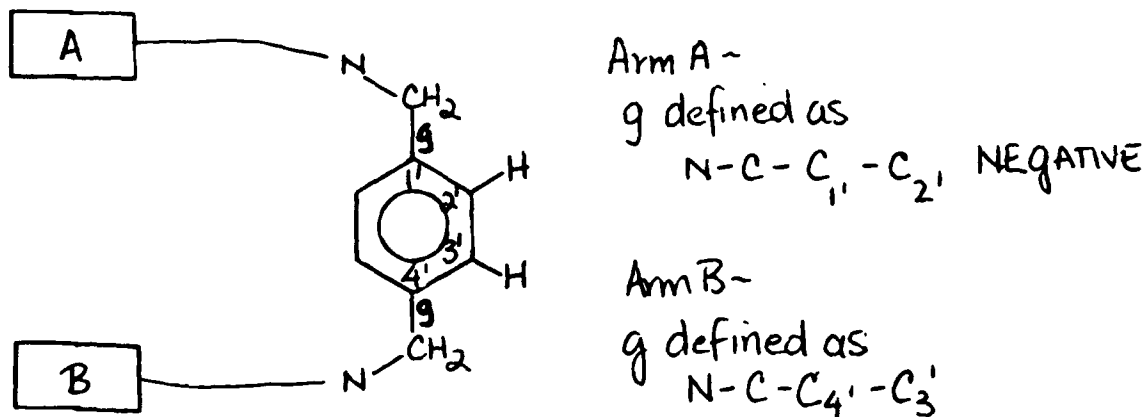
The p-xylylene aromatic spacer provides a more symmetrical clam shape due to the para substitution of the steroidal arms onto the benzene ring. In addition, the N-H to C_2-H interaction is not as severe. Therefore, since the considerations of geometry were different here, the studies with this host proceeded differently than with the m-xylylene fragment. Two combinations of dihedral angles **b** and **g** giving folded "C" shapes were studied, along with two extended "S" shapes. The starting structures in this series were optimized energetically (full geometry, no electrostatics) using the MAXIMIN2 force field found in the SYBYL Molecular Modeling Software (Appendix A). The resulting "C" shapes widened in the middle while moving closer together at the ends, the A rings of the steroidal arms. The "S" shapes tightened in their shape by curving more toward the spacer. The following table shows both the beginning and final energies and dihedral angles for each of the p-xylylene cholic acid hosts.

Name	Energy	Dihedral Angles for Arm A , Arm B	
		Bond 20-22 (b)	Bond g
_PX	+817	+60.6, +60.6	-90, +90
_PX_MAX[a]	+65.1	+62.6, +62.7	-82.6, +97
_PX_0	+1028	+60.6, 0	-90, +90
_PX_0_MAX	+71.06*	+59.9, -35.4	-73.3, +105
_PX_OPEN[b]	+817	+60.6, +60.6	-90, -90
_PX_OPEN_MAX	+67.16	+63.5, +63.7	-85.5, -85.7
_PX_0E[c]	+1029	+60.6, 0	-90, -90
_PX_0E_MAX	+74.54**	+56.6, -38.0	-79.5, -71.8

* +0.04 away from convergence at +71.023
** +0.04 away from convergence at +74.40

[a] _MAX = MAXIMIN2 result, all _MAX results were fit to _PX

[b] the "open" or extended form of _PX
c the extended form of _PX_0

The components of the total energy are given in Appendix B. The initial difference between the two 90 degree clam shapes is shown in Fig. 7. Initial and final structures _PX_0 and _PX_0_MAX are shown in Fig. 8, illustrating the typical change in the "C" clam shape. The structure of _PX_MAX is shown in Fig. 9. Finally, Fig. 10 shows how minimization alters the "S" shape of an extended p-xylylene cholic acid host.

The atom pair distances indicative of the cavities created in the two "C" shapes (both before and after minimization) were:

Name	Å Distance Arm A // B	O3	O7	O12	Cavity Width (Å)
	O3	10.00	12.72	9.34	
_PX	O7	12.67	14.72	11.29	8.81
	O12	9.26	11.27	8.81	
	A // B	O3	O7	O12	
	O3	9.98	12.68	9.12	
_PX_MAX	O7	12.63	14.86	11.41	9.08
	O12	9.04	11.40	9.08	
	A // B	O3	O7	O12	
	O3	3.01	6.54	8.50	
_PX_0	O7	7.48	8.28	10.57	6.47, 10.57*
	O12	7.78	6.47	9.63	
	A // B	O3	O7	O12	
	O3	2.90	6.56	8.32	
_PX_0_MAX	O7	7.31	9.20	10.85	6.66, 10.85
	O12	7.78	6.47	9.63	

*cavity is larger in the middle than at the ends

The two 90 degree clam shape cavities formed from the p-xylylene aromatic spacer, with widths of 9-11 Å and lengths of 12 Å, are reasonable starting points for incorporating the monosaccharide moiety. The energy minimizations did not significantly alter the cavity sizes but optimized the overall geometry (Fig. 8). The extended forms of these hosts are created by rotation about a singe bond, **g** in arm **B**, from the flexible chain to the aromatic ring. If the molecule exists in an extended form, for example conformation _PX_OPEN_MAX, the distance from the end of the steroid arm **B** to the plane of the xylylyl fragment is 12.03 Å (Fig. 11). Therefore, to create a clam or "C" shape, this arm would have to swing around on the hinge to cover almost 25Å!

To complete this phase of the cholic acid host system geometry evaluation, energy minimizations on the m-xylylene structures must be done and the accompanying change in cavity size and/or shape must be studied. Docking a monosaccharide guest, n-pentyl beta-D-glucoside, has begun in the p-xylylene cholic acid hosts by first visually docking the sugar and then optimizing the whole complex using MAXIMIN2. These studies are preliminary since only the steric (no electrostatic) component to the energy has been included in the evaluation of the host guest complexes.
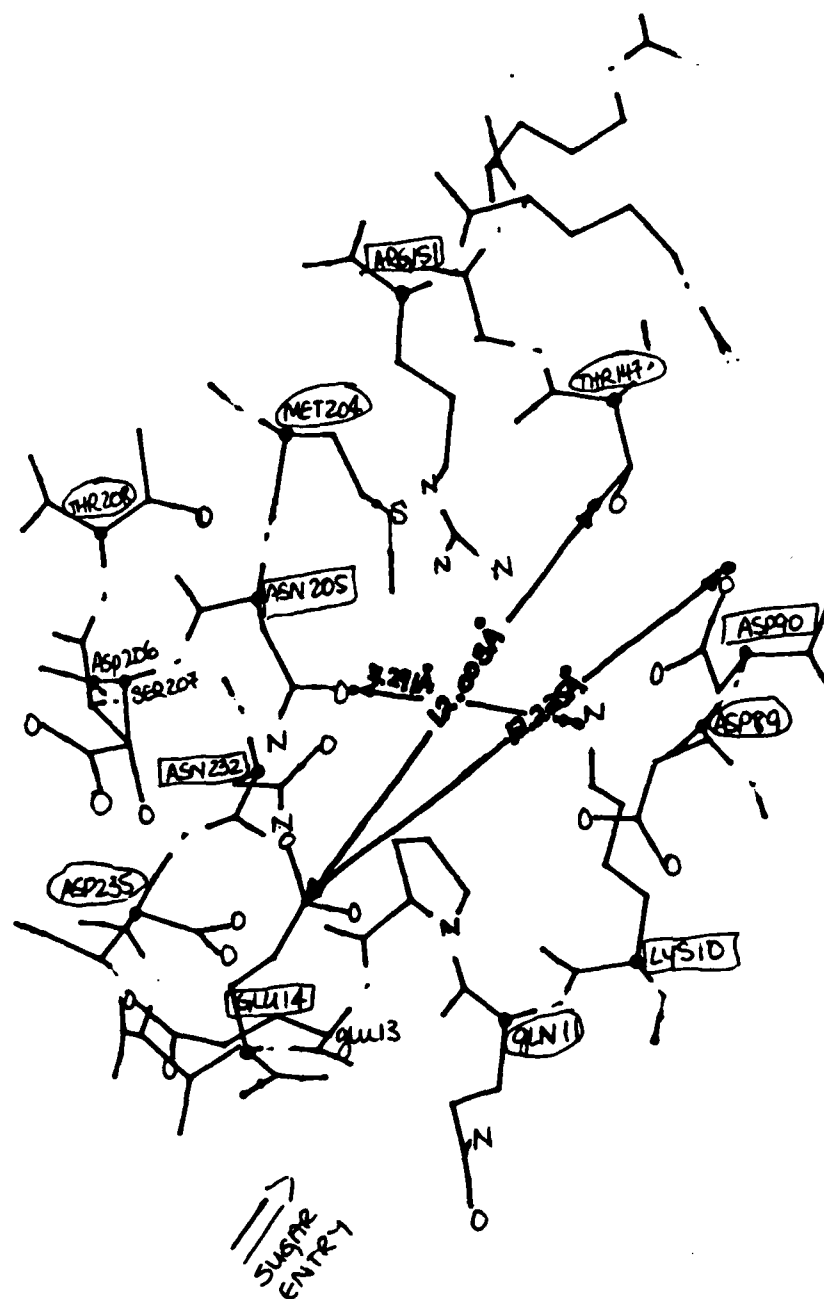
FIGURE 1.
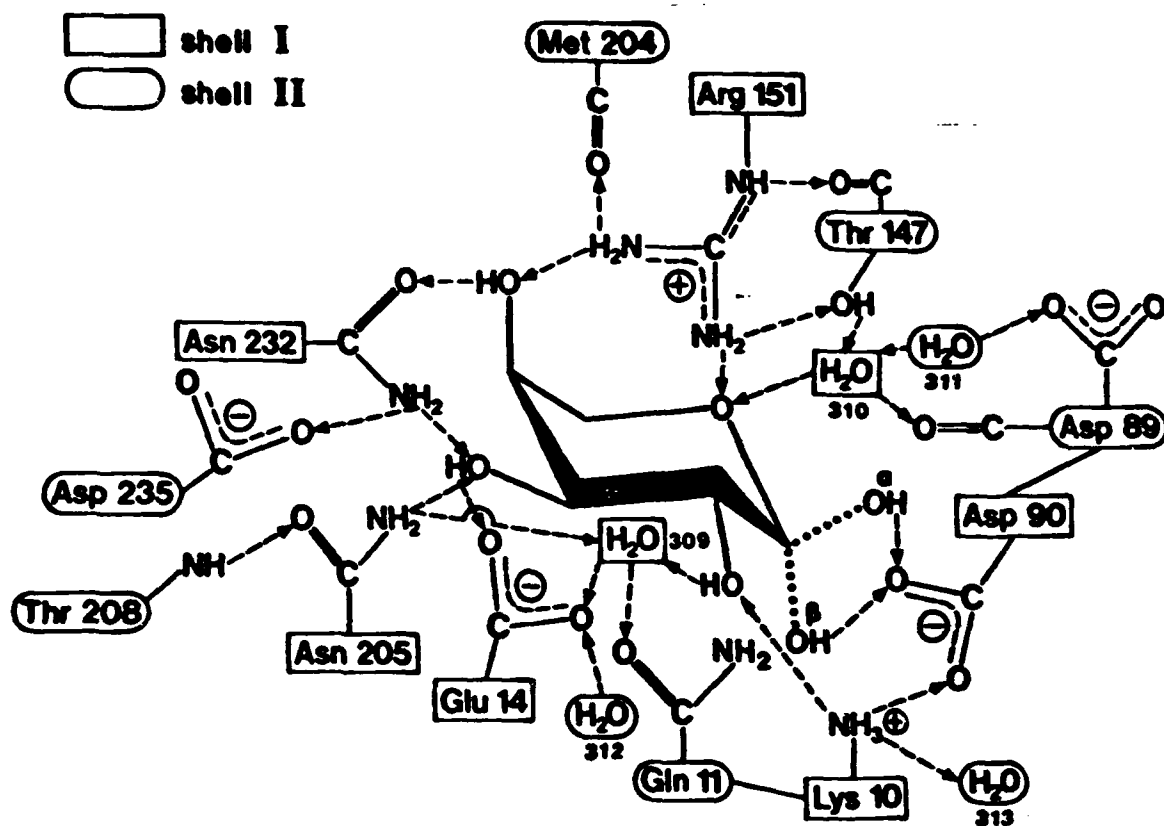
L_ABP_12345 BY_ATOM_COLOR   NOV 08 1989

Figure 2. Schematic diagram of the intricate networks of hydrogen bonds formed in the complex of the L-arabinose-binding protein with the L-arabinose substrate. Shell I represents the essential residues hydrogen-bonded to the sugars and to adjacent second shell (shell II) of residues. Note especially that Arg-151 is not involved in salt linkages but is the source of five donor groups for five hydrogen bonds. (Adapted from Quiocho and Vyas 1984.)
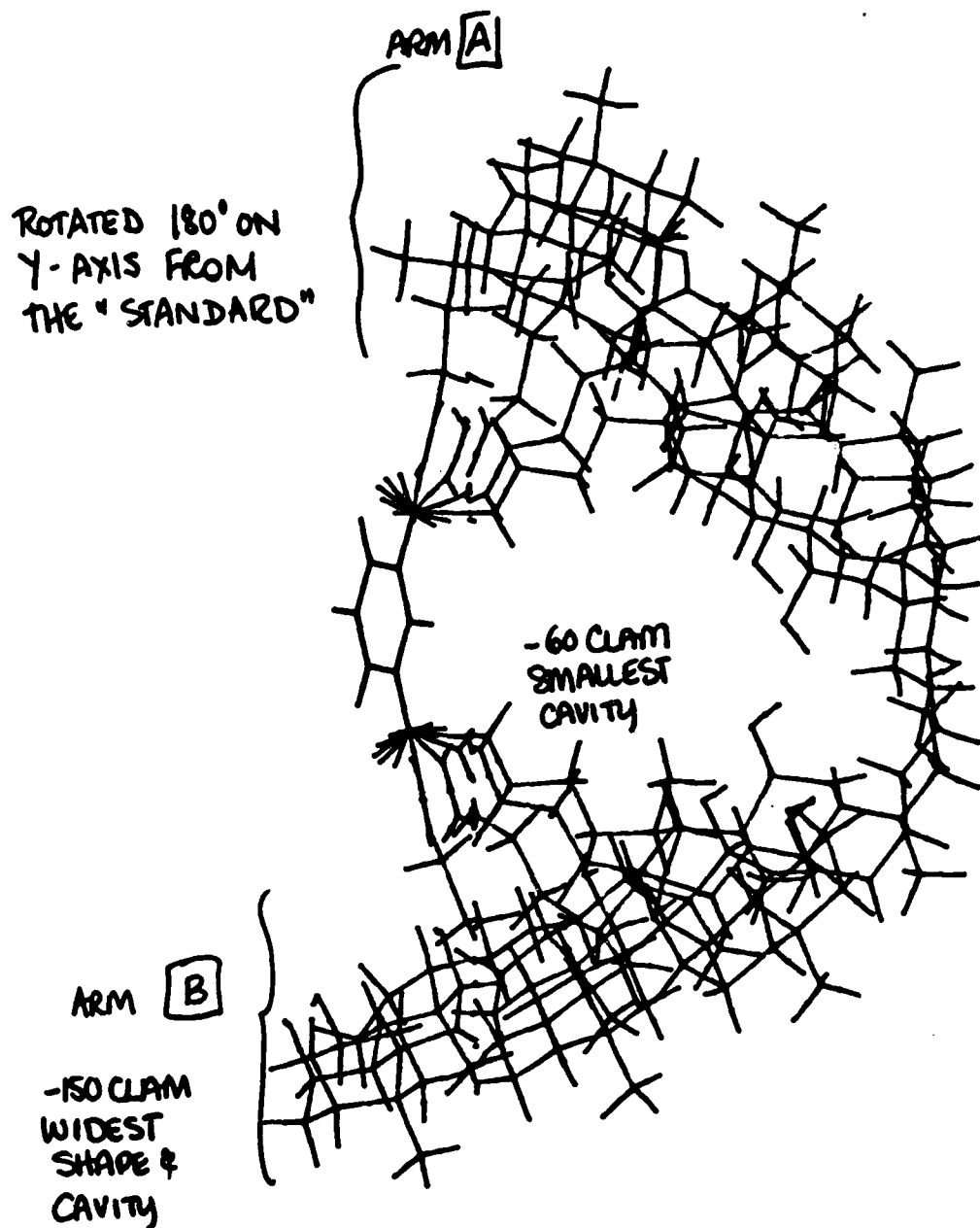
FIGURE 2.

ARM [A]

ROTATED 180° ON
Y·AXIS FROM
THE "STANDARD"

~60 CLAM
SMALLEST
CAVITY

ARM [B]

~150 CLAM
WIDEST
SHAPE &
CAVITY

FIGURE 3 .

ALL BUGJES_MX2_* FITTED

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TOTAL OF 5 STRUCTURES

IN THE "STANDARD" ORIENTATION, OH'S DIRECTED OUTWARD, NOT INWARD INTO CAVITY
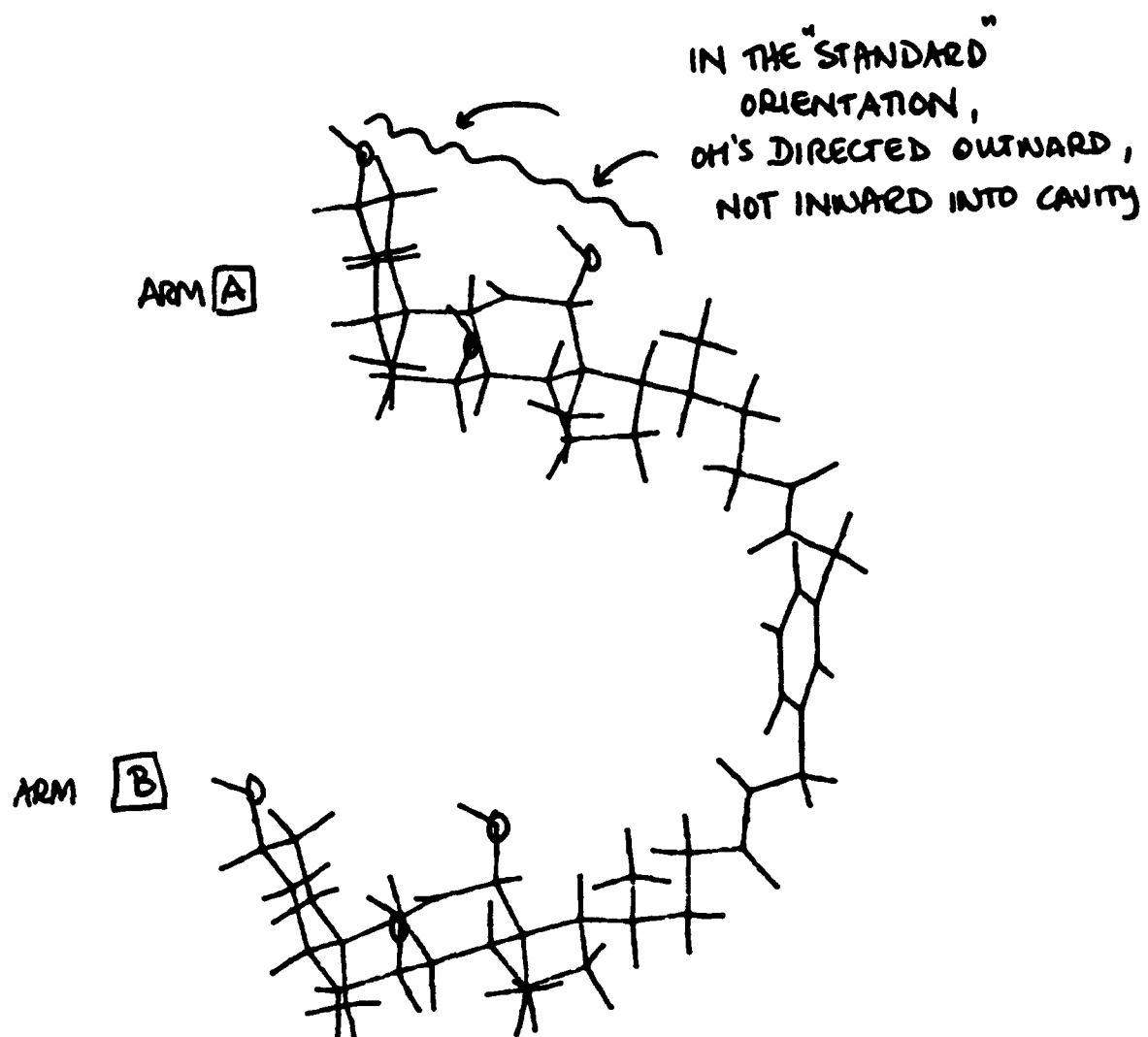
ARM A

ARM B

FIGURE 4.

BUGJES_MX2_90-b

IN "STANDARD"
ORIENTATION

ARM A)

_90

_90E
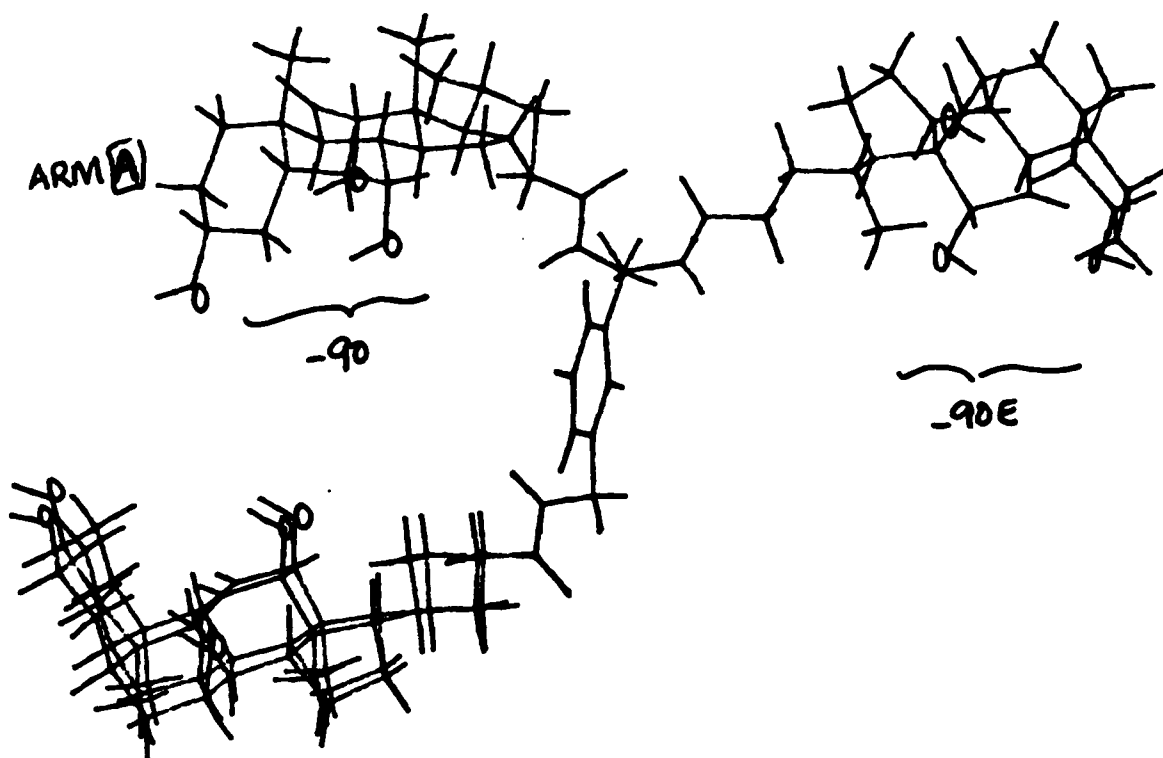
FIGURE 5.

BUGJES_MX2_90E (EAST) ON _90

APR 1988

_MI
ARM Ⓐ

_MX2_150
ARMⒶ

_MI
ARM Ⓑ

_150
Ⓑ
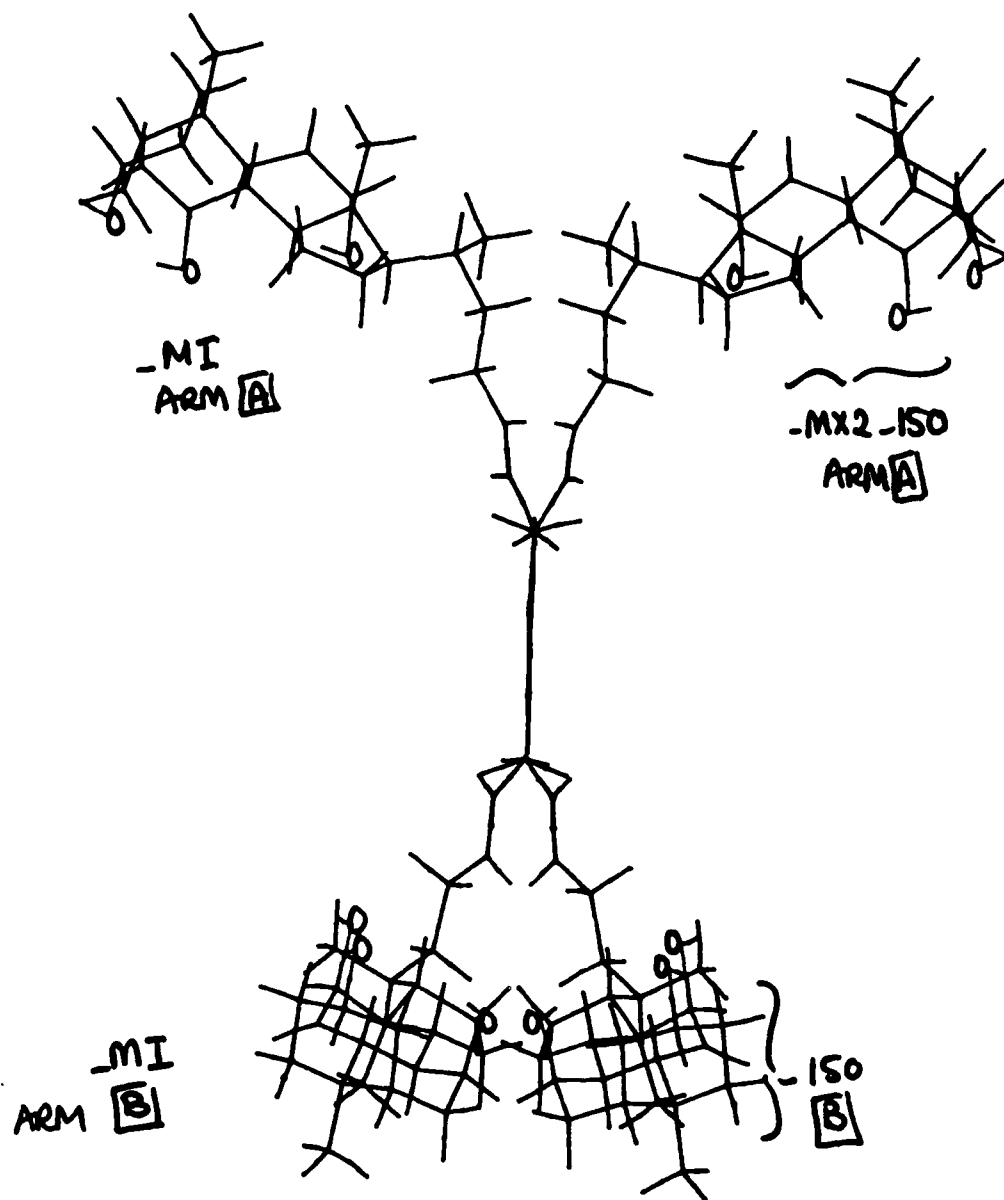
FIGURE 6.

BUGJES_MX2_150 EAST ON _MX2_MI WEST

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IN "STANDARD" ORIENTATION

ARM A

-PX-φ ---->

-PX ---->
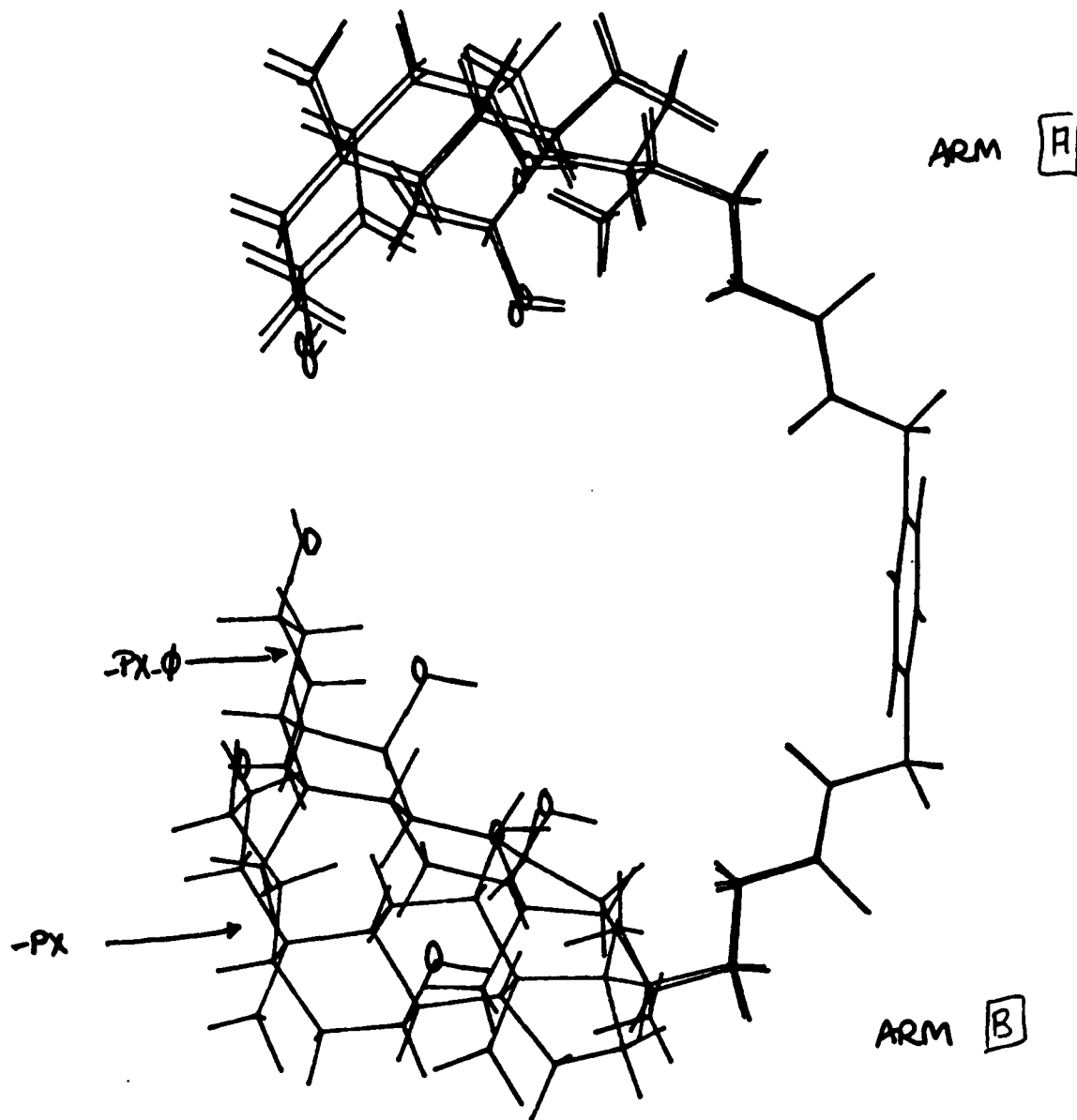
ARM B

FIGURE 7.

CUBSUN_PX & CUBSUN_PX_0 (RED) ARM A -->ARM B

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_PX_φ_MAX

SMALLER

SLIGHTLY WIDER
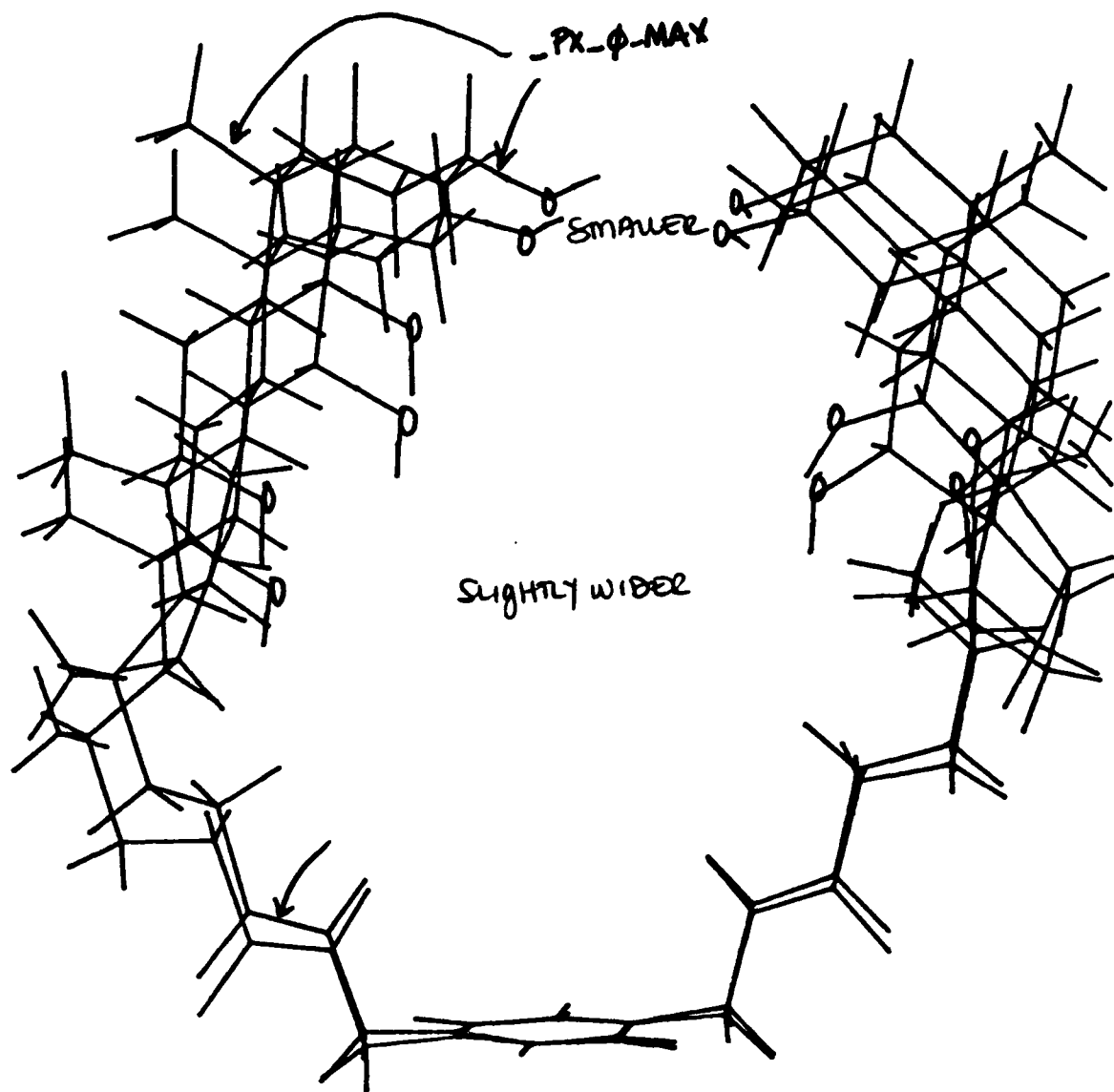
FIGURE 8.

CUBSUN_PX_φ_MAX (RED) ON _PX_φ

APK
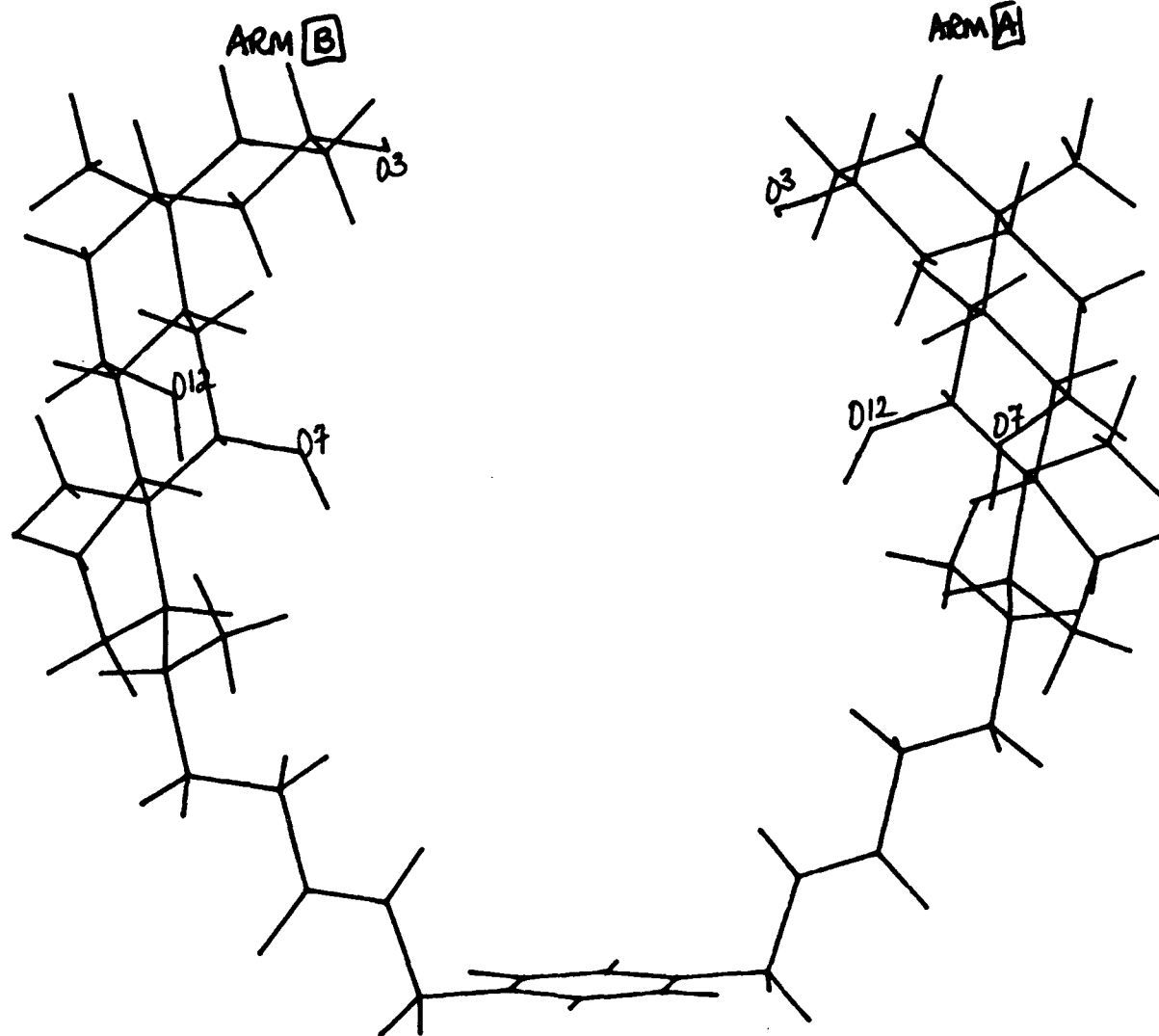
FIGURE 9.

CUBSUN_PX_MAX

1APR

ARM A

_PX.OPEN_MAX

ARM B
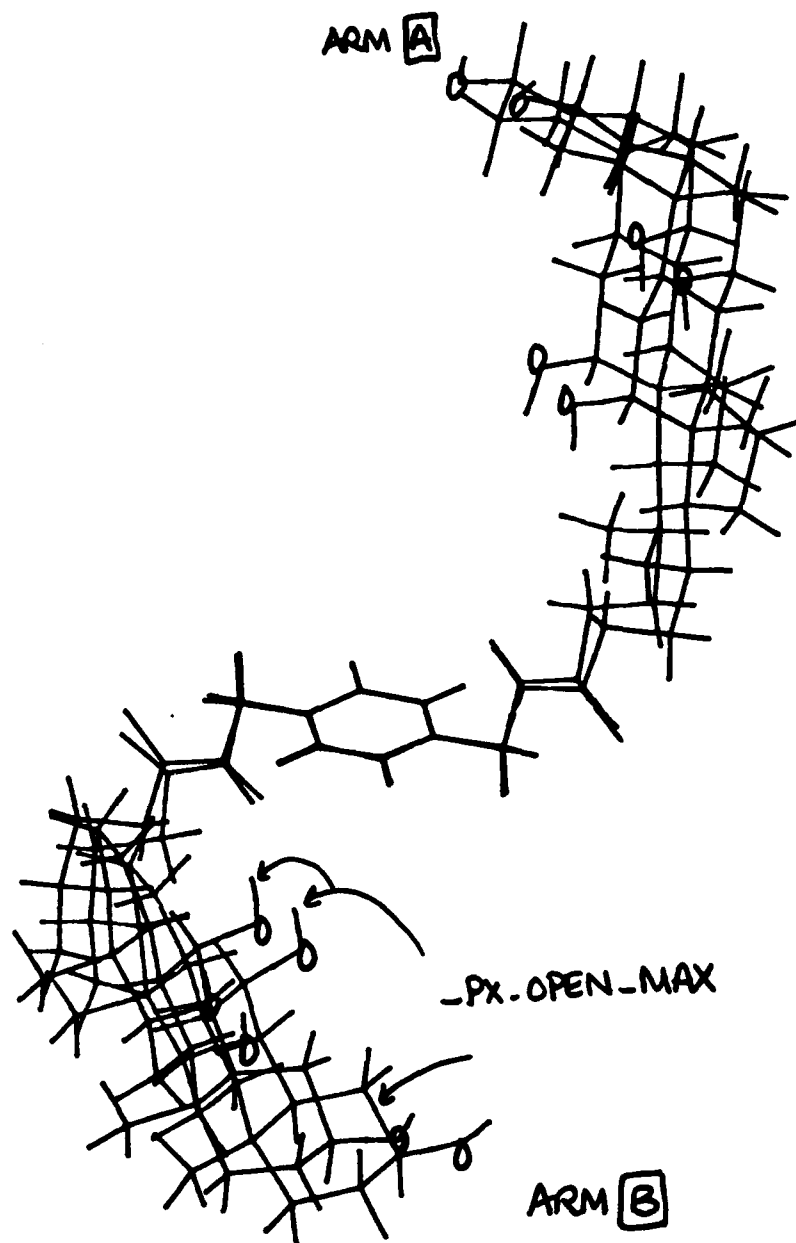
Figure 10.

CUBSUN_PX_OPEN_MAX ON GREEN INPUT

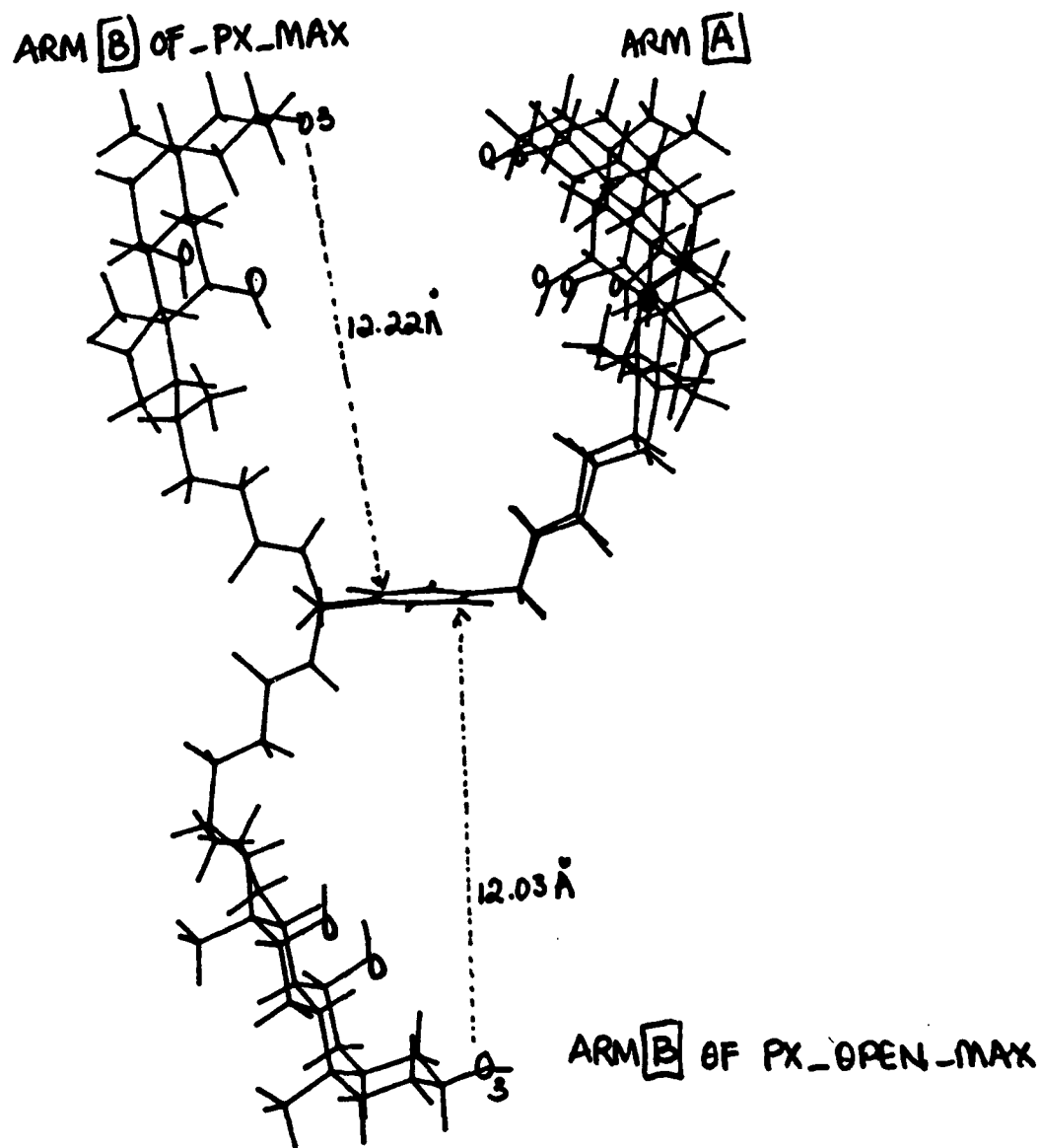APR 00 1000

ARM B OF_PX_MAX

ARM A

12.22Å

12.03Å

ARM B OF PX_OPEN_MAX

FIGURE 11.

CUBSUN_PX_MAX (BL) & CUBSUN_PX_OPEN_MAX